Total Synthesis of Seco (+)- and *ent*-(-)-Oxaduocarmycin SA: **Construction of the (Chloromethyl)indoline Alkylating Subunit by** a Novel Intramolecular Aryl Radical Cyclization onto a Vinyl Chloride

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A practical, total synthesis of seco-(+)-oxaduocarmycin **3a**, an analogue of the highly cytotoxic natural product, duocarmycin SA (1), is described. The 13-step synthesis features a novel and efficient intramolecular aryl radical cyclization onto a vinyl chloride as a direct entry to the (chloromethyl)indoline alkylating subunit 14. Subsequent resolution, utilizing a preparative Chiralpak AD column, provided enantiomerically pure alkylating subunits 14a and 14b which were elaborated to seco-(+) and *ent*-(-)-oxaduocarmycins, **3a** and **3b**, respectively. The natural enantiomer 3a was active at pM concentrations and exhibited 7–50-fold higher potentcy than its enantiomer 3b in in vitro cytotoxicity assays.

Introduction

(+)-Duocarmycin SA (1),² isolated from *Streptomyces* sp. DO113, is the newest member of a class of potent antitumor antibiotics^{3,4} that are structurally and biologically related to (+)-CC-1065 (2).5 In common, these natural products possess a cyclopropanoindolinone linked to a DNA binding subunit as the pharmacophore responsible for the sequence-selective minor groove alkylation of duplex DNA⁵ and the exceptionally potent cytotoxic activity.^{5,6} Detailed studies, however, reveal two critical features that distinguish (+)-duocarmycin SA (1) in its interaction with duplex DNA and suggest that it may

represent the most exciting agent in the series.^{7,8} The first feature relates to the extent of the noncovalent DNA binding⁸ which stabilizes the *reversible*⁹ alkylation of DNA while providing (+)-duocarmycin SA (1) with the full biological properties of the irreversible alkylating agent, (+)-CC-1065 (2).¹⁰ This, in combination with the implications that the extent of the noncovalent stabilization may be related to the delayed, fatal hepatotoxicity associated with (+)-CC-1065 (2),9a,10 suggests that (+)duocarmycin SA (1) may prove to be a better therapeutic agent. Second, as the most stable natural product in this class of agents,¹¹ (+)-duocarmycin SA^{12} (1) is also the most potent,^{6a} a finding that is consistent with a correlation between solvolytic stability and cytotoxic potency that was established within a series of agents possessing sufficient reactivity to alkylate DNA.¹³ Together, these characteristics provide rationale for the development of (+)-duocarmycin SA (1) and related agents as potent oncolytics for the treatment of human cancers.¹⁴

(12) The name duocarmycin SA originates from duocarmycin stable

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⁽²⁾ Isolation: (a) Ichimura, M.; Ogawa, T.; Takahashi, K.; Koba-yashi, E.; Kawamoto, I.; Yasuzawa, T.; Takahashi, I.; Nakano, H. *J. Antibiot.* **1990**, *43*, 1037. (b) Ichimura, M.; Ogawa, T.; Katsumata, S.; Takahashi, K.; Takahashi, I; Nakano, H. J. Antibiot. 1991, 44, 1045. (c) Yasuzawa, T.; Muroi, K.; Ichumura, M.; Takahashi, I.; Ogawa, T.; Takahashi, K.; Sano, H.; Saitoh, Y. Chem. Pharm. Bull. 1995, 43, 378. Total syntheses: (d) See ref 30. (e) Boger, D. L.; Machiya, K.; Hertzog, D. L.; Kitos, P. A.; Holmes, D. J. Am. Chem. Soc. 1993, 115, 9025. (f) Muratake, H.; Abe, I.; Natsume M. Chem. Pharm. Bull. 1996, 44, 67. (g) Murake, H.; Tonegawa, M.; Natsume, M. Chem. Pharm. Bull. 1996, *4*4. 1631.

⁽³⁾ Duocarmycin A, B₁, B₂, C₁, and C₂: (a) Ichimura, M; Muroi, K.;
Asano, K.; Kawamoto, I.; Tomita, F.; Morimoto, M.; Nakano, H. J. Antibiot. **1988**, 41, 1285. (b) Takehashi, I.; Takehashi, K.; Ichimura, M.; Morimoto, M.; Asano, K.; Kawamoto, I.; Tomita, F.; Nakano, H. J. Antibiot. 1988, 41, 1915. (c) Yasuzawa, T.; Iida, T.; Muroi, K.; Ichimura, M.; Takahashi, K.; Sano, H. *Chem. Pharm. Bull.* **1988**, *36*, 3728. (d) Ogawa, T.; Ichimura, M.; Katsumata, S.; Morimoto, M.; Takahashi, K. J. J. Antibiot. 1989, 42, 1299.

<sup>K. J. J. Antibiot. 1989, 42, 1299.
(4) Pyrindamycin A and B: (a) Ohba, K.; Watabe, H.; Sasaki, T.; Takeuchi, Y.; Kodama, S. J. Antibiot. 1988, 41, 1515. (b) Ishii, S.; Nagasawa, M.; Kariya, Y.; Yamamoto, H.; Inouye, S.; Kondo, S. J. Antibiot. 1989, 42, 1713.
(5) (a) Hurley, L. H.; Needham-VanDevanter, D. R. Acc. Chem. Res. 1986, 19, 230. (b) Warpehoski, M. A.; Hurley, L. H. Chem. Res. Toxicol. 1988, 1, 315. (c) Coleman, R. S.; Boger, D. L. In Studies in Natural Products Chemistry Atta-ur-Rahman Ed.: Elsevier: Amsterdam. 1989;</sup>

Products Chemistry, Atta-ur-Rahman, Ed.; Elsevier: Amsterdam, 1989; Vol. 3, p 301. (d) Boger, D. L. In Advances in Heterocyclic Natural Products Synthesis; Pearson, W. H., Ed.; JAI Press: Greenwich, CT, 1992; Vol. 2, pp 1–188. (e) Boger, D. L. Acc. Chem. Res. 1995, 28, 20 (f) Boger, D. L.; Johnson, D. S. Angew. Chem., Int. Ed. Engl. 1996, 35, 1439

^{(6) (}a) Boger, D. L.; Ishizaki, T.; Zarrinmayeh, H.; Munk, S. A.; Kitos, P. A.; Suntornwat, O. J. Am. Chem. Soc. 1990, 112, 8961. (b) Sugiyama, H.; Hosoda, M., Saito, I. Tetrahedron Lett. 1990, 31, 7197. (c) Warpehoski, M. A. In *Advances in DNA Sequence Specific Agents*; Hurley, L. H., Ed.; JAI: Greenwich, CT, 1992; Vol. 1, p 217. (d) Hurley, L. H.; Draves, P. H. In *Molecular Aspects of Anticancer Drug–DNA Interac-tions*; Neidle, S., Waring, M., Eds; CRC: Ann Arbor, MI, 1993; Vol. 1, p 89.

⁽⁷⁾ Boger, D. L. In Advances in Nitrogen Heterocycles; Moody, C. J.,

<sup>Ed.; JAI Press: Greenwich, CT, 1995; Vol. 1, pp 229–247.
(8) (a) Boger, D. L.; Johnson, D. S.; Yun, W. J. Am. Chem. Soc. 1994, 116, 1635. (b) Boger, D. L.; Johnson, D. S. Proc. Natl. Acad. Sci. U.S.A.</sup> 1995, 92, 3642. (c) Boger, D. L.; Johnson, D. S. J. Am. Chem. Soc. 1995, 117 1443

^{(9) (}a) Boger, D. L.; Yun, W. J. Am. Chem. Soc. **1993** 115, 9872. (b) Asai, A.; Nagamura, S.; Saito, H.; Takahashi, I; Nakano, H. Nucleic Acids Res. 1994, 22, 88.

⁽¹⁰⁾ Warpehoski, M. A.; Harper, D. E.; Mitchell, M. A.; Monroe, T.

 ⁽¹⁰⁾ Walpenski, N. A., Halper, D. E., Mitchell, M. T., Marten, M. T., Marten, J. Biochemistry 1992, *31*, 2502.
 (11) (a) Boger, D. L.; Coleman, R. S.; Invergo, B. J.; Sakya, S. M.; Ishizaki, T.; Munk, S. A.; Zarrinmayeh, H.; Kitos, P. A.; Thompson, S. C. *J. Am. Chem. Soc.* 1990, *112*, 4623. (b) Boger, D. L. In *Heterocycles* in Bioorganic Chemistry; Bergman, J., van der Plas, H. C. Simonyi, M., Eds.; Royal Society of Chemistry: Cambridge, 1991. (c) Boger, D. L. In Proc. Robert A. Welch Found. Conf. Chem. Res. 35th Chem. Front. Med. 1991. 35. 137.

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Chart 1





3a: seco-(+)-Oxa-Duocarmycin SA

Seco-(+)-oxaduocarmycin SA **3a** (LY307918) is an analogue in which the fused pyrrole ring in the alkylating subunit of **1** is replaced with a potentially metabolically more inert furan ring,¹⁵ and the cyclopropanoindolinone pharmacophore is masked as a (chloromethyl)hydroxy-indoline, a prodrug form which is known to demonstrate indistinguishable biological properties from its active form.¹⁶ The unexpected observation that both (+)-**1** and its enantiomer *ent*-(-)-**1** alkylate DNA with comparable efficiency,¹⁷ prompted us to evaluate both (+)-**3a** and *ent*-(-)-**3b** as potentially useful oncolytics. A practical synthetic route was, therefore, required to provide sufficient quantities of both the enantiomers for a full evaluation of their biological properties.

Herein, we report an efficient and practical total synthesis of (+)-**3a** and *ent*-(-)-**3b** in which the key transformation involves a C–C bond formation via a *novel 5-exo-trig cyclization of an aryl radical onto a vinyl chloride*¹⁸ to provide a direct entry into the (chlorometh-yl)indoline ring system (Scheme 1).¹⁹ Subsequent resolution, by chromatographic separation on a chiral column, leads to enantiomerically pure alkylating subunits, which are then elaborated to furnish both enantiomers of seco-oxaduocarmycin SA (**3**).²⁰



Results and Discussion

Alkylation of the potassium salt of commercially available hydroxybenzaldehyde **4** with ethyl bromoacetate cleanly provided 5^{21} (Scheme 2). Intramolecular aldol cyclization of **5** followed by dehydration was most effectively achieved in one step with DBU in refluxing ethanol to directly give benzofuran 6^{21} in a modest 44% yield. O-Demethylation, with concomitant ester hydrolysis, occurred on heating **6** with excess, neat pyridine hydrochloride²² at 170 °C to give hydroxy acid **7**, which was directly reesterified with acidic methanol to give the desired methyl ester **8** in an 95% overall yield. Reduction

^{(13) (}a) Boger, D. L.; Ishizaki, T. Tetrahedron Lett. 1990, 31, 793.
(b) Boger, D. L.; Munk, S. A.; Ishizaki, T. J. Am. Chem. Soc. 1991, 113, 2779.
(c) Ichimura, M.; Ogawa, T.; Takasahi, K.; Mihara, A.; Takahashi, I.; Nakano, H. Oncol. Res. 1993, 5, 165.
(d) Boger, D. L.; Goldberg, J.; McKie, J. A. Bioorg. Med. Chem. Lett. 1996, 6, 1955.

⁽¹⁴⁾ KW-2189, a novel derivative of duocarmycin B2, is presently in clinical trials: (a) Asai, A.; Nagamura, S.; Saito, H. J. Am. Chem. Soc. **1994**, *116*, 4171. (b) Kobayashi, E.; Okamato, A.; Asada, M.; Okabe, M.; Nagamura, S.; Asai, A.; Saito, H.; Gomi, K.; Hirata, T. Cancer Res. **1994**, *54*, 2404. (c) Nagamura, S.; Asai, A.; Kanda, Y.; Kobayashi, E.; Gomi, K.; Saito, H. Chem. Pharm. Bull. **1996**, *44*, 1723.

⁽¹⁵⁾ Mohammadi, F.; Spees, M. M.; Staten, G. S.; Marder, P.; Kipka, J. K.; Johnson, D. A.; Boger, D. L.; Zarrinmayeh, H. *J. Med. Chem.* **1994**, *37*, 232.

⁽¹⁶⁾ Seco-(+)-duocarmycin SA exhibits cytotoxic activity as well as a DNA alkylating selectivity and efficiency identical with those of (+)duocarmycin SA: see ref 8a.

⁽¹⁷⁾ *ent*-(–)-Duocarmycin SA alkylates DNA but only at concentrations approximately $10 \times$ that required for (+)-duocarmycin SA. Consequently, the natural enantiomer is $10 \times$ more effective than the unnatural enantiomer: see ref 5e.

⁽¹⁸⁾ To our knowledge, the *intramolecular* addition of carbon radicals to vinyl chlorides has not been reported; however, vinyl fluorides are known to be suitable radical acceptors: (a) Morikawa, T.; Uchida, J.; Hasegawa, Y.; Taguchi, T. *Chem. Pharm. Bull.* **1991**, *39*, 2464. (b) Morikawa, T.; Uchida, J.; Imoto, K.; Taguchi, T. *J. Fluorine Chem.* **1992**, *58*, 119. (c) Dolbier, W. R., Jr.; Rong, X. X. Tetrahedron Lett. **1996**, *37*, 5321. The corresponding *intermolecular* reactions with vinyl chlorides and vinyl fluorides are well-documented: (d) Low, H. C.; Tedder, J. M.; Walton, J. C. J. Chem. Soc., *Faraday Trans.* **1976**, *72*, 1707. (e) Henning, R.; Urbach, H. *Tetrahedron Lett.* **1983**, *24*, 5343. (f) Giese, B.; Gonzalez-Gomez, J. A.; Witzel, T. Angew. Chem., Int. Ed. Engl. **1984**, *23*, 69.

⁽²⁰⁾ This approach provides a useful improvement to established synthetic approaches to such agents: (a) 5-exo-dig aryl radical cyclization: Boger, D. L.; Ishizaki, T.; Wysocki, R. J., Jr.; Munk, S. A.; Kitos, P. A.; Suntornwat, O. J. Am. Chem. Soc. **1989**, 111, 6461. Boger, D. L.; Ishizaki, T.; Kitos, P. A.; Suntornwat, O. J. Org. Chem. Soc. **1990**, 55, 5823. (b) 5-exo-trig aryl radical-alkene cyclization (functionalized alkene): Boger, D. L.; Yun, W.; Teegarden, B. R. J. Org. Chem. Soc. **1992**, 57, 2873. (c) Boger, D. L.; Mckie, J. A. J. Org. Chem. **1995**, 60, 1271.

^{(21) (}a) Aneja, R.; Mukerjee, S. K.; Seshadri, T. R. *Tetrahedron* **1958**, *2*, 203. (b) Sahm, W.; Schinzel, E.; Jurges, P. *Justus Liebigs Ann. Chem.* **1974**, 523. (c) Gammill, R. B.; Nash, S. A. *J. Org. Chem.* **1986**, *51*, 3116.



of the nitro group in **8**, using H_2 over 5% Pd-Al₂O₃ in a solvent mixture of THF:MeOH, followed by protection of the resulting amine 9, led to Boc aniline 10. O-Benzylation of phenol 10. effected under standard Mitsunobu²³ conditions, then gave fully protected benzofuran 11. A low-temperature, electrophilic aromatic bromination of 11, using NBS in the presence of catalytic concentrated H₂SO₄,²⁴ led to regiospecific introduction of the C-4 bromide to provide 12 as the sole product in an excellent yield. Deprotonation of amide 12, using NaH, followed by alkylation²⁴ of the resulting anion with commercially available (E:Z)-1,3-dichloropropene gave an inconsequential mixture of E:Z isomers of vinyl chloride 13, the desired precursor for the key aryl radical cyclization (Scheme 3). A deoxygenated solution of aryl bromide 13 in 0.015 M benzene was heated at reflux for 3.5 h in the presence of 1.1 equiv of tri-n-butyltin hydride and cat. AIBN to give the desired, fully functionalized alkylating subunit 14 in 72% yield.²⁵ Particularly noteworthy was the simple workup employed that involved trituation in hexanes followed by filtration to provide pure 14 as a white solid with no detectable levels of tin residues. The cyclized product 14 is formed in a highly chemoselective manner believed to proceed via an initial, preferential homolysis of the weaker aryl C-Br bond²⁶ in **13** to generate an aryl radical which undergoes a preferred



5-exo-trig intramolecular cyclization²⁷ onto the proximal vinyl chloride acceptor. A hydrogen radical quench of the resulting (chloromethyl)indoline radical terminates the chain reaction. The absence of a methylindoline product that could originate from reduction of **14** indicates that the chloromethyl functionality was, indeed, stable to homolysis under these reaction conditions.

Resolution,²⁸ on a preparatively useful scale was readily accomplished by direct chromatographic separation of racemate **14** on a preparative Chiralpak AD (Amylose)²⁹ column (8 cm × 24 cm, n-PrOH:hexane (51: 49), column volume (CV) = 964 mL, 225 mL/min flow rate), to provide multigram quantities (360 mg in 300 mL eluent/injection) of enantiomers (+)-**14a** ($t_{\rm R}$ = 5.6 min, 97.6 % ee, 49 % recovery) and *ent*-(-)-**14b** ($t_{\rm R}$ = 7.4 min, 95.55 % ee, 48 % recovery). Acid-mediated deprotection

⁽²²⁾ Rene L.; Buisson J–P.; Roger, R. Bull. Soc. Chim. Fr. 1975, 11–12, 2763.

⁽²³⁾ Mitsunobu, O. Synthesis 1981, 1.

 ⁽²⁴⁾ Boger, D. L.; Coleman, R. S. J. Am. Chem. Soc. 1988, 110, 1321.
 (25) A minor uncyclized product (<2%), resulting from reduction of the aryl bromide, was detected by ¹H NMR but not isolated.
 (26) An automatic of metric of a constraint of a constraint of the aryle bromide.

⁽²⁶⁾ An example of regiospecific homolysis of a C-Br bond in the presence of a C-Cl bond has been reported: Stork, G.; Mook, R. J. Am. Chem. Soc. **1983**, 105, 3721.

^{(27) (}a) Baldwin, J. E. J. Chem. Soc., Chem. Commun. 1976, 734. (b) Beckwith, A. J. L.; Schiesser, C. H. Tetrahedron 1985, 41, 3945.

⁽²⁸⁾ The amine, derived from deprotection of 14, proved unstable and was unamenable to resolution through classical fractional recrystallization of diastereomeric salts. Covalent diastereomers, prepared by attachment of chiral agents to either the amine or phenol derivatives of 14, were not readily separable by chromatograhic means.

⁽²⁹⁾ Effective separations on related agents have been previously achieved by direct chromatographic resolution on a ChiraCel OD column: (a) Boger, D. L.; Yu, W. J. Am. Chem. Soc. **1994**, 116, 7996. (b) Boger, D. L.; Bollinger, B.; Johnson, D. S. *Bioorg. Med. Chem. Lett.* **1996**, 6, 2207. (c) Boger, D. L. et al, J. Am. Chem. Soc. **1997**, 119, 4977. See also ref 8a.

Synthesis of Seco (+)- and *ent*-(-)-Oxaduocarmycin SA

Table 1. In Vitro Cytotoxicity, IC₅₀ (nM)

	tumor cell line		
	T222	CCRF-CEM	GC3/C1
(+)- 3a ent-(-)- 3b	0.39 19.4	0.23 1.59	0.10 0.70

of (+)-14a and *ent*-(-)-14b selectively removed the Boc group to give unstable amine salts (+)-15a and *ent*-(-)-15b, which were directly coupled with 5,6,7-trimethoxy-indole-2-carboxylic acid^{6a} 16, under standard EDCI-promoted coupling conditions,³⁰ to furnish enantiomerically pure penultimate intermediates (+)-17a (96 % ee) and *ent*-(-)-17b (94 % ee), respectively, in overall 50% yield. Final, biphasic, transfer catalytic hydrogenolysis³¹ of 17a and 17b served to quantitatively remove the benzyl ether and provided (+)-3a ($[\alpha]^{23}_{D} - 1.73^{\circ}$ (*c* 0.11, DMF))³² and its enantiomer (-)-3b ($[\alpha]^{23}_{D} - 2.1^{\circ}$ (*c* 0.12, DMF)), respectively.

The biological activity, determined in an *in vitro* cytotoxicity assay,³³ showed that duocarmycin SA analogue, (+)-**3a** (GC3/C1; IC₅₀ = 100 pM) was highly potent and in the same range as that of the natural product (+)-**1** (L1210; IC₅₀ = 10 pM). Furthermore, the natural enantiomer (+)-**3a** displayed 7–50-fold higher potentcy than the unnatural isomer, *ent*-(-)-**3b** (Table 1), consistent with the biological findings of duocarmycin SA ((+)-1) and its enantiomer.^{8a} These preliminary results are sufficiently encouraging to warrant further studies to fully characterize the biological properties of these agents and provided the incentive to develop the refined synthetic approach detailed herein.

Conclusion

A concise, total synthesis of seco-(+)- and *ent*-(-)oxaduocarmycins has been achieved in 13 synthetic steps. Central to this strategy was an aryl radical cyclization onto a vinyl chloride, followed by a preparatively useful resolution procedure. The tactical improvement defined in this work shortens approaches to related agents²⁰ by 2-4 synthetic steps. This methodology should find application as a general synthetic approach to other members of this class of agents and related analogues.

Experimental Section

Ethyl 2-(6-Formyl-2-methoxy-4-nitrophenoxy)acetate (5). To a rapidly stirring solution of 2-hydroxy-3-methoxy-5nitrobenzaldehyde 4 (100 g, 0.507 mol) in dry methanol (1.5 L) was added powdered potassium hydroxide (85%, 37.1 g, 0.562 mol) and the mixture heated at reflux for 45 min. Methanol solvent was removed in vacuo and the resulting solid residue suspended in dimethylformamide (1.5 L). The mixture was cooled to 0 °C and ethyl bromoacetate (103 mL, 0.925 mol) added. The solution was warmed to room temperature and stirred for 36 h. The solvent was removed in vacuo and the resulting solid residue was diluted with water (3 L) and extracted with ethyl acetate (8 \times 1 L). The combined organic extracts were washed with water (1 L) and brine (1 L), dried (Na₂SO₄), and concentrated to give the desired product 5 (136.5 g, 95%) as a tan solid: mp 109-110 °C; ¹H NMR (DMSO-d₆, 300 MHz) δ 10.48 (s, 1H), 8.09 (d, J = 2.94 Hz, 1H), 8.06 (d,

(33) Procedures are described in the experimental section. For T222 tumor cell line also see: ref 15; for CCRF–CEM and GC3/C1 tumor cell lines also see: Mosmann, T. J. Immunol. Meth. **1983**, 65, 55–62.

Methyl 7-Methoxy-5-nitrobenzofuran-2-carboxylate (6). 1,8-Diazabicyclo[5.4.0]undec-7-ene (80 mL, 0.535 mol) was added to a solution of benzaldehyde 5 (136.4 g, 0.482 mol) in ethanol (2.5 L) and the solution heated at reflux for 4 h. The reaction mixture was cooled to room temperature and the precipitated solid filtered, washed with cold ethanol (600 mL), and dried (in vacuo at 40 °C) to provide benzofuran 6 (56.1 g, 44%) as a yellow solid: mp 161–162 °C; ¹H NMR (DMSO- d_6 , 250 MHz) δ 8.38 (d, J = 1.84 Hz, 1H), 7.93 (s, 1H), 7.90 (d, J= 2.21 Hz, 1H), 4.40 (q, J = 6.99 Hz, 2H), 4.10 (s, 3H), 1.36 (t, J = 6.99 Hz, 3H); ¹³C NMR (CDCl₃, 63 MHz) δ 158.3 (C), 148.4 (C), 147.8 (C), 145.8 (C), 145.3 (C), 127.7 (C), 114.2 (CH), 111.3 (CH), 103.8 (CH), 61.9 (CH₂), 56.5 (CH₃), 14.2 (CH₃); IR (KBr) $v_{\rm max}$ 3500, 3200, 3000, 2950, 1718, 1529, 1345, 1322, 1297, 1188, 1096, 979, 944, 882, 739 cm⁻¹; UV (EtOH) λ_{max} 213 ($\epsilon =$ 14538), 264.5 (ϵ = 28797) nm; FDMS m/z 265 (M⁺, 100). Anal. Calcd for C₁₂H₁₁NO₆: C, 54.34; H, 4.18; N, 5.28. Found: C, 54.08; H, 4.10; N, 5.29.

7-Hydroxy-5-nitrobenzofuran-2-carboxylic Acid (7). Methoxybenzofuran 6 (55.9 g, 0.211 mol) was combined with pyridine hydrochloride (243 g, 2.11 mol) in a flask equipped with a paddle stirrer. The two solids were heated neat at 170 °C with vigorous stirring for 18 h. The mixture was allowed to cool to \sim 50 °C and then poured into ice-water (2 L). The precipitated solid was filtered and washed with water (1 L). The crude solid was dissolved in 1 N sodium hydroxide (1.5 L) and washed with ethyl acetate (2×500 mL). Concentrated hydrochloric acid was added to the aqueous solution until pH 3 was reached. The precipitated product was filtered, washed with water, and dried (in vacuo at 50 °C) to give phenol 7 (44 g, 88%) as a pale yellow solid: mp 189-191 °C; ¹H NMR (DMSO- d_6 , 300 MHz) δ 13.93 (s, 1H), 11.41 (s, 1H), 8.21 (d, J = 2.21 Hz, 1H), 7.81 (s, 1H), 7.71 (d, J = 2.21 Hz, 1H); IR (KBr) v_{max} 3274 (br), 3093, 2542, 1694, 1530, 1347, 1287, 1225, 1193, 791 cm⁻¹; UV (EtOH) λ_{max} 208 ($\epsilon = 15442$), 263.5 ($\epsilon =$ 18538) nm; FDMS m/z 223 (M⁺, 100). Anal. Calcd for C₉H₅NO₆: C, 48.44; H, 2.26; N, 6.28. Found: C, 48.74; H, 2.41; N, 6.28.

Methyl 7-Hydroxy-5-nitrobenzofuran-2-carboxylate (8). Acetyl chloride (13 mL, 186 mmol) was added dropwise to dry methanol (1.5 L) at 0 °C and the solution stirred at room temperature for 0.5 h. Carboxylic acid 7 (39.6 g, 177 mmol) was added to the acidic methanol and the mixture heated at reflux, under a nitrogen atmosphere, for 18 h. The reaction was cooled to room temperature and the resulting precipitate filtered, washed with cold methanol (500 mL), and dried to give methyl ester 8 (41.2 g, 98%) as a light tan solid: mp 151-152 °C; ¹H NMR (DMSO- d_6 , 300 MHz) δ 11.48 (s, 1H), 8.23 (d, J = 2.21 Hz, 1H), 7.91 (s, 1H), 7.73 (s, 1H), 3.93 (s, 3H); IR (KBr) v_{max} 3281, 1688, 1582, 1523, 1442, 1356, 1328, 1256, 1102, 969, 901, 810, 744 cm⁻¹; UV (EtOH) λ_{max} 213 (ϵ = 16556), 267.5 (ϵ = 26855) nm; FDMS *m*/*z* 238 (M⁺, 100). Anal. Calcd for C₁₀H₇NO₆: C, 50.64; H, 2.97; N, 5.91. Found: C, 50.85; H, 3.01; N, 5.88.

Methyl 5-Amino-7-hydroxybenzofuran-2-carboxylate (9). To a solution of nitrobenzofuran 8 (42 g, 177 mmol) in tetrahydrofuran:methanol (1:1, 1.5 L) was added 5% palladium-Al₂O₃ (5 g) and the mixture stirred at room temperature under 60 psi hydrogen atmosphere for 4 h. The reaction mixture was filtered through Celite and the filtrate concentrated *in vacuo* to give a solid, which was purified by flash chromatography (gradient: 0-5% methanol:dichloromethane) to provide aminobenzofuran 9 (35.6 g, 97%) as a tan solid: mp $201-202 \,^{\circ}$ C; ¹H NMR (DMSO- d_6 , 250 MHz) δ 9.93 (s, 1H), 7.44 (s, 1H), 6.29 (d, J = 2 Hz, 1H), 6.24 (d, J = 1.75 Hz, 1H), 4.92 (s, 2H), 3.85 (s, 3H); IR (KBr) v_{max} 3416, 3335, 2947, 1713,

⁽³⁰⁾ Boger, D. L.; Machiya, K. J. Am. Chem. Soc. 1992, 114, 10056.
(31) (a) Ram, S.; Ehrenkaufer, R. E. Synthesis 1988, 91. Bieg, T.;
Szeja, W. Synthesis 1985, 76.

1610, 1568, 1454, 1434, 1356, 1204, 1155 cm⁻¹; UV (EtOH) λ_{max} 235.5 (ϵ = 20635), 290.5 (ϵ = 15655) nm; FDMS *m*/*z* 207 (M⁺, 100). Anal. Calcd for C₁₀H₉NO₄ requires: C, 57.97; H, 4.38; N, 6.76%. Found: C, 57.72; H, 4.28; N, 6.63%.

Methyl 5-(N-(tert-Butyloxycarbonyl)amino)-7-hydroxybenzofuran-2-carboxylate (10). To a solution of amine 9 (35.6 g, 172 mmol) in tetrahydrofuran (750 mL) was added di-tert-butyl dicarbonate (48 g, 222 mmol) and the mixture stirred at room temperature for 24 h. The solvent was removed in-vacuo and the residue dissolved in ethyl acetate (1.5 L), washed with 1 N aqueous hydrochloric acid (2 \times 200 mL), water (2 \times 600 mL), and brine (1 \times 600 mL), and then dried (Na₂SO₄). The organic solution was passed through a pad of silica gel and concentrated in vacuo to furnish Boc amine **10** (49.4 g, 93%) as a white powder: mp 190–192 °C; ¹H NMR (DMSO- d_6 , 300 MHz) δ 10.34 (s, 1H), 9.34 (s, 1H), 7.65 (s, 1H), 7.37 (d, J = 1.47 Hz, 1H), 7.10 (d, J = 1.84 Hz, 1H), 3.88 (s, 3H), 1.48 (s, 9H); IR (KBr) v_{max} 3291, 1698, 1606, 1584, 1534, 1453, 1340, 1249, 1164, 1109, 1062, 976, 852, 763, 739 cm⁻¹; UV (EtOH) λ_{max} 211 (ϵ = 25983), 244.5 (ϵ = 31990), 287.5 ($\epsilon = 19236$) nm; FDMS m/z 308 (M⁺, 100). Anal. Calcd for C₁₅H₁₇NO₆: C, 58.63; H, 5.58; N, 4.56. Found: C, 58.88; H, 5.63; N, 4.60.

Methyl 5-(N-(tert-Butyloxycarbonyl)amino)-7-(benzyloxy)benzofuran-2-carboxylate (11). To a solution of phenol 10 (48.9 g, 159 mmol) in dichloromethane (1.5 L) were added benzyl alcohol (30 mL, 291 mmol) and triphenylphosphine (75.1 g, 286 mmol) at room temperture and under a dry nitrogen atmosphere. The solution was cooled to 0 °C and diethyl azodicarboxylate (DEAD) (50 g, 287 mmol) added dropwise. The reaction mixture was stirred at room temperature for 18 h and concentrated in vacuo and the solid residue purified by flash chromatoghaphy (gradient: 5-20% ethyl acetate/hexanes) to provide benzyl ether 11 (57 g, 90%) as a white solid: mp 141–143 °C; ¹H NMR (DMSO- d_6 , 250 MHz) δ 9.43 (s, 1H), 7.71 (s, 1H), 7.55 (s, 2H), 7.53 (d, J = 1.75 Hz, 1H), 7.51-7.39 (m, 3H), 7.29 (d, J = 1.75 Hz, 1H), 5.22 (s, 2H), 3.87 (s, 3H), 1.48 (s, 9H); 13 C NMR (CDCl₃, 63 MHz) δ 159.6 (C), 152.9 (C), 146.0 (C), 144.6 (C), 142.1 (C), 136.1 (C), 135.2 (C), 128.6 (C), 128.5 (CH), 128.0 (CH), 127.6 (CH), 114.2 (CH), 104.3 (CH), 103.7 (CH), 80.5 (C), 71.0 (CH₂), 52.1 (CH₃), 28.3 (3 x CH₃); IR (KBr) v_{max} 3440, 2982, 2954, 2934, 1725, 1606, 1582, 1528, 1320, 1158 cm⁻¹; UV (EtOH) λ_{max} 242 (ϵ = 29172), 280 (ϵ = 16141) nm; FDMS *m*/*z* 397 (M⁺, 100). Anal. Calcd for C₂₂H₂₃NO₆: C, 66.48; H, 5.83; N, 3.52. Found: C, 66.25; H, 5.86; N, 3.69.

Methyl 5-(N-(tert-Butyloxycarbonyl)amino)-7-(benzyloxy)-4-bromobenzofuran-2-carboxylate (12). A solution of benzofuran 11 (30 g, 75.5 mmol) in dry tetrahydrofuran (250 mL) was cooled to -60 °C and two drops of concentrated sulfuric acid, followed by N-bromosuccinamide (14.8 g, 83 mmol), added under a stream of dry nitrogen. The mixture was warmed to 0 °C over a period of 3 h and was then quenched with 0.1 N aqueous sodium bisulfite (5 mL). The organic solvent was removed in vacuo and the residue diluted with ethyl acetate (300 mL), washed with water (2×150 mL), and brine (2 \times 150 mL), dried (Na₂SO₄), and concentrated in vacuo to afford aryl bromide $\mathbf{12}$ (34.3 g, 95%) as a light yellow solid: mp 140–142 °C; ¹H NMR (DMSO- d_6 , 300 MHz) δ 8.07 (s, 1H), 7.54-7.49 (m, 3H), 7.42-7.34 (m, 3H), 6.94 (s, 1H), 5.31 (s, 2H), 3.97 (s, 3H), 1.55 (s, 9H); 13C NMR (CDCl₃, 63 MHz) & 159.2 (C), 152.5 (C), 146.0 (C), 144.2 (C), 141.2 (C),135.8 (C),133.2 (C), 129.1 (C), 128.4 (CH), 128.2 (CH), 128.0 (CH), 114.0 (CH), 104.6 (CH), 94.2 (C), 81.1 (C), 71.2 (CH₂), 52.3 (CH₃), 28.2 (3 \times CH₃); IR (KBr) v_{max} 3700, 3440, 3000, 2983, 17231453, 1233, 1157 cm⁻¹; UV (EtOH) λ_{max} 212.5 ($\epsilon =$ 27223), 246 (ϵ = 28581), 287 (ϵ = 14281) nm; FDMS *m*/*z* 475 (M⁺, 100). Anal. Calcd for C₂₂H₂₂NO₆Br: C, 55.48; H, 4.65; N, 2.94; Br, 16.78%. Found: C, 55.20; H, 4.70; N, 3.20; Br, 16.50

Methyl 5-[*N*-(*tert*-Butyloxycarbonyl)-*N*-(3-chloro-2propen-1-yl)amino]-7-(benzyloxy)-4-bromobenzofuran-2carboxylate (13). Boc amine 12 (1.43 g, 3 mmol) was added portionwise to a suspension of sodium hydride (60% in mineral oil; 156 mg, 3.90 mmol) in dry dimethylformamide (8.0 mL) at room temperature and under nitrogen. The resulting dark yellow mixture was stirred at room temperature for 40 min and cooled to 0 °C and then neat (E:Z)-1,3-dichloropropene (846 μ L, 9 mmol) added. The reaction was allowed to warm to room temperature, stirred for a further 15 h, quenched with brine (4 mL), and extracted with ethyl acetate (3 \times 150 mL). Combined, dried (MgSO₄) organics were concentrated in vacuo, and the resulting crude brown oil was purified by flash chromatography (gradient: 0-20% ethyl acetate/hexanes) to give vinyl chloride 13 (1.36 g, 82%) as a low melting yellow glass: (E:Z vinyl chlorides and NBoc rotomers): ¹H NMR (CDCl₃, 300 MHz) δ 7.56 (s, 1H), 7.48–7.20 (m, 5H), 6.85– 6.71 (m, 1H), 5.99-5.85 (m, 2H), 5.32 (br s, 2H), 4.46 (dd, J= 15.5 and 6.2 Hz, 1H), 4.35-4.20 (m, 1H), 3.98 (s, 3H), 1.5 (br s, 3H), 1.28 (br s, 6H);¹³C NMR (CDCl₃, 63 MHz) δ 159.1 (C), 153.8 (C), 146.4 (C), 146.3 (C), 135.8 (C), 135.7 (C), 128.6 (CH), 128.5 (CH), 128.3 (CH), 128.2 (CH), 127.4 (CH), 127.0 (CH), 120.7 (CH), 114.8 (CH), 114.7 (C), 113.3 (CH), 80.6 (C), 71.4 (CH₂), 71.3 (CH₂), 52.4 (CH₃), 28.0 (CH₃); IR (CHCl₃) v_{max} 2981, 1729,1 698, 1368, 1165 cm⁻¹; UV (EtOH) λ_{max} 203 (ϵ = 32586), 245.5 (ϵ = 30885), 286.5 (ϵ = 17684) nm; FDMS *m*/*z* 551 (M⁺, 100). Anal. Calcd for C₂₅H₂₅NO₆Cl Br: C, 54.51; H, 4.57; N, 2.58; Cl, 6.44; Br, 14.51; Found: C, 54.31; H, 4.60; N, 2.68; Cl, 6.72; Br, 14.42.

Methyl (1R/S)-5-(Benzyloxy)-3-(tert-butyloxycarbonyl)-1-(chloromethyl)-1,2-dihydro-3H-furano[3,2-e]indole-7carboxylate (14). To a solution of aryl bromide 13 (12.30 g, 22.34 mmol) in dry benzene (1.49 L) were added tri-n-butyltin hydride (6.52 mL, 24.57 mmol) and catalytic 2,2'-azobis(isobutyronitrile) (AIBN) (183 mg, 1.12 mmol). The solution was deoxygenated by bubbling dry nitrogen through the solution at room temperature for ~ 1 h. The reaction mixture was then heated at reflux for 3.5 h and concentrated in vacuo to give an oily residue. Trituation of the crude oil with hexanes provided a solid which was filtered and washed with hexanes (3 \times 500 mL) to give indoline 14 (7.79 g, 72%) as an off-white solid: mp 137-139 °C; ¹H NMR (CDCl₃, 300 MHz) δ 7.85 (br s, 1H), 7.54–7.30 (m, 6H), 5.28 (s, 2H), 4.17 (apparent t, $J\!=\!$ 10.4 Hz, 1H), 4.08-3.98 (m, 1H), 3.95 (s, 3H), 3.91-3.71 (m, 2H), 3.56 (apparent t, J = 10.3 Hz,1H), 1.56 (s, 9H); ¹³C NMR (CDCl₃, 63 MHz) δ 159.4 (C), 152.2 (C), 146.5 (C), 144.9 (C), 142.3 (C), 136 1 (C), 128.4 (CH), 128.0 (CH), 127.7 (CH), 111.6 (CH), 100.1 (C), 70.9 (CH₂), 52.9 (CH₂), 52.2 (CH₃), 46.6 (CH₃), 41.5 (C), 28.4 (3 \times CH₃); IR (CHCl₃) v_{max} 2956, 1725, 1695, 1497, 1158 cm⁻¹; UV (EtOH) λ_{max} 219 (ϵ = 27621), 254.5 (ϵ = 38111), 285 (ϵ = 14659), 346 (ϵ = 2729) nm; FDMS *m*/*z* 471 (M⁺, 100). Anal. Calcd for C₂₅H₂₆NO₆Cl; C, 63.63; H, 5.55; N, 2.97; Cl, 7.51; Found: C, 63.35; H, 5.51; N, 2.74; Cl, 7.30.

Resolution of 14. Twenty micron Chiralpak AD (Amylose) (600 g) was slurried in 1-propanol (1200 mL) and transferred to a 8 cm Prochrom column and the bed compressed at a pressure of 85 Bar resulting in a 8 cm \times 24 cm configuration with a column volume (CV) of 964 mL. Preparative separations were preformed at a flow rate of 225 mL/min with a column pressure of 11 Bar. Racemate **14** (360 mg) was dissolved in eluent 1-propanol/hexanes (51:49, 300 mL) and charged on the column using a sample pump. Chromatograms from a run on the Prochrom column is illustrated in Figure 1. Racemate **14** (10.57 g) was resolved, in 35 runs on the 8 cm column, with an optimized time of 13 min per run and effluent monitored at 280 nm, to give:

Methyl (1.5)-5-(Benzyloxy)-3-(*tert*-butyloxycarbonyl)-1-(chloromethyl)-1,2-dihydro-3*H*-furano[3,2-*e*]indole-7carboxylate (14a). First peak ($t_{\rm R} = 5.6$ min, 97.6 % ee, 5.16 g, 49 % recovery), as a white solid: $[\alpha]^{23}_{\rm D} - 10.8^{\circ}$ (*c* 0.5, CHCl₃). Mp 130–132 °C dec; ¹H NMR (CDCl₃, 300 MHz) δ 7.90 (br s, 1H), 7.55–7.33 (m, 6H), 5.29 (s, 2H), 4.18 (apparent dd, J =11.3 and 10 Hz, 1H), 4.08–3.93 (m, 1H), 3.96 (s, 3H), 3.87– 3.80 (m, 2Hl), 3.60 (apparent t, J = 10.4 Hz, 1H), 1.57 (s, 9H); ¹³C NMR (CDCl₃, 63 MHz) δ 159.4 (C), 152.2 (C), 146.5 (C), 144.9 (C), 142.3 (C), 136 1 (C), 128.4 (CH), 128.0 (CH), 127.7 (CH), 111.6 (CH), 100.1 (C), 70.9 (CH₂), 52.9 (CH₂), 52.2 (CH₃), 46.6 (CH₃), 41.5 (C), 28.4 (3 × CH₃); IR (KBr) $v_{\rm max}$ 2990, 1737, 1696, 1152, 1137, 767, 696 cm⁻¹; UV (EtOH) $\lambda_{\rm max}$ 219 ($\epsilon =$ 27870), 254.5 ($\epsilon =$ 39132), 284.5 ($\epsilon =$ 14997), 346 ($\epsilon =$ 2901) nm; FDMS *m*/*z* 470.9 (M⁺, 100). Anal. Calcd for C₂₅H₂₆NO₆-



Figure 1. Typical chromatograms of the separation of racemate **14** on a Chiralpak AD (Amylose) column.

Cl: C, 63.63; H, 5.55; N, 2.97; Cl, 7.51; Found: C, 63.56; H, 5.53; N, 2.67; Cl, 7.72.

Methyl (1R)-5-(Benzyloxy)-3-(tert-butyloxycarbonyl)-1-(chloromethyl)-1,2-dihydro-3H-furano[3,2-e]indole-7carboxylate (14b). Second peak ($t_{\rm R} = 7.4 \text{ min}, 95.5 \%$ ee, 5.04 g, 48 % recovery) as a white solid: $[\alpha]^{23}_D$ +11.2° (*c* 0.5, CHCl₃). mp 131–133 °C dec; ¹H NMR (CDCl₃, 300 MHz) δ 7.90 (br s, 1H), 7.55-7.28 (m, 6H), 5.29 (s, 2H), 4.18 (dd, J = 11.6and 10 Hz,1H), 4.08-3.98 (m, 1H), 3.96 (s, 3H), 3.88-3.80 (m, 2Hl), 3.60 (apparent t, J = 10.4 Hz, 1H), 1.57 (s, 9H); ¹³C NMR (CDCl₃, 63 MHz) & 159.4 (C), 152.2 (C), 146.5 (C), 144.9 (C), 142.3 (C), 136 1 (C), 128.4 (CH), 128.0 (CH), 127.7 (CH), 111.6 (CH), 100.1 (C), 70.9 (CH₂), 52.9 (CH₂), 52.2 (CH₃), 46.6 (CH₃), 41.5 (C), 28.4 (3 \times CH₃); IR (KBr) $v_{\rm max}$ 2990, 1736, 1697, 1499, 1424, 1349, 1208, 1152, 1137, 762, 697 cm⁻¹; UV (EtOH) λ_{max} 219.5 (ϵ = 26377), 254.5 (ϵ = 37628), 284.5 (ϵ = 14440), 346.5 $(\epsilon = 2823)$ nm; FDMS m/z 470.9 (M⁺, 100). Anal. Calcd for C₂₅H₂₆NO₆Cl: C, 63.63; H, 5.55; N, 2.97; Cl, 7.51; Found: C, 63.91; H, 5.58; N, 3.13; Cl, 7.59.

Methyl (15)-5-(Benzyloxy)-1-(chloromethyl)-1,2-dihydro-3H-furano[3,2-e]indole-7-carboxylate (15a). Boc indoline 14a (800 mg, 1.697 mmol) was added to a dry solution of 4 N HCl in dioxane (25 mL) at 0 °C and under nitrogen. The solution was stirred at 0 °C for 8 h and the solvent removed at 0 °C under high vacuum to provide amine hydrochloride salt 15a (690mg, 100%) as an unstable brown solid: ¹H NMR (DMSO-d₆, 300 MHz) δ 8.07 (s, 1H), 7.55-7.52 (m, 2H), 7.47-7.39 (m, 3H), 7.13 (s, 1H), 5.32 (s, 2H), 4.14-3.91 (m, 2H), 3.91 (s, 3H), 3.73-3.60 (m, 2H), 3.51-3.47 (m, 1H); IR (KBr) v_{max} 3400 (br), 1717, 1584, 1333, 1317, 1207, 1141, 1104, 915, 727 cm⁻¹; UV (EtOH) λ_{max} 243 (ϵ = 18536), 285 (ϵ = 12985), 359 (ϵ = 2272), 389 (ϵ = 1674) nm; FDMS m/z 371 (Free amine, M⁺, 100). Anal. Calcd for C₂₀H₁₉NO₄Cl₂: C, 58.84; H, 4.69; N, 3.43; Cl, 17.37. Found: C, 58.71; H, 4.85; N, 3.13; Cl, 16.82

Methyl (1*R***)-5-(Benzyloxy)-1-(chloromethyl)-1,2-dihydro-3***H***-furano[3,2-***e***]indole-7-carboxylate (15b). Boc indoline 14b** (1.00g, 2.12 mmol) was added to a dry solution of 4 N HCl in dioxane (28 mL) at 0 °C and under nitrogen. The solution was stirred at 0 °C for 8 h and the solvent removed at 0 °C under high vacuum to provide amine hydrochloride salt **15a** (865mg, 100%) as an unstable brown solid: ¹H NMR (DMSO-*d*₆, 300 MHz) δ 8.08 (s, 1H), 7.55–7.52 (m, 2H), 7.47– 7.39 (m, 3H), 7.15 (s, 1H), 5.32 (s, 2H), 4.14–3.91 (m, 2H), 3.91 (s, 3H), 3.72–3.61 (m, 2H), 3.51–3.48 (m, 1H); IR (KBr) v_{max} 3400 (br), 1717, 1583, 1333, 1317, 1275, 1207, 1141, 1104, 728 cm⁻¹; UV (EtOH) λ_{max} 211 (ϵ = 26721), 243 (ϵ = 18796), 285 (ϵ = 13166), 359 (ϵ = 2189) nm; FDMS *m*/*z* 371 (Free amine, M⁺, 100). Anal. Calcd for C₂₀H₁₉NO₄Cl₂: C, 58.84; H, 4.69; N, 3.43; Cl, 17.37. Found: C, 58.78; H, 4.72; N, 3.18; (Cl, 16.82).

Methyl (1*R*)-5-(Benzyloxy)-1-(chloromethyl)-1,2-dihydro-3-[(5,6,7-trimethoxy-1*H*-indol-2-yl)carbonyl]-3*H*-furano[3,2-*e*]indole-7-carboxylate (17a). To a dry solution of amine hydrochloride salt 15a (173 mg, 0.424 mmol) in dimethylformamide (4.0 mL) was added sodium bicarbonate (178 mg, 2.12 mmol) followed by 5,6,7-trimethoxyindolecarboxylic acid 16 (128 mg, 0.509 mmol) and finally 1-(3-(dimethylamino)propyl)-3-ethylcarbodiimide hydrochloride (EDCI) (244 mg, 1.272 mmol) and the mixture stirred at room temperature under nitrogen for 18 h. The solvent was reduced to $\sim 1/2$ the original volume and ethyl acetate (20 mL) added. The organics were washed with 0.1 N aqueous hydrochloric acid (2×10 mL), water (10 mL), saturated sodium bicarbonate (10 mL), and brine (10 mL) and then dried (MgSO₄). Concentration of the organics followed by purification of the resulting crude solid by flash chromatography (gradient: 10-40% ethyl acetate/ hexanes) provided amide 17a (127 mg, 50%, HPLC (analytical ChiraCel OD column; flow rate = 1.0 mL/min; UV detection at $\lambda = 254$ nm; eluent, CH₃CN:H₂O 90:10): $t_{\rm R} = 11.08$ min; 96 % ee) as a white powder: $[\alpha]^{23}_{D} + 13.6^{\circ}$ (*c* 0.5, CHCl₃). mp 182-183.5 °C (ÉtOAc/hexanes, white needles); ¹H NMR (CDCl₃, 300 MHz) & 9.38 (brs, 1H), 8.31 (s, 1H), 7.56-7.52 (m, 3H), 7.43–7.28 (m, 3H), 6.96 (d, J = 2.1 Hz, 1H), 6.87 (s, 1H), 5.35 (s, 2H), 4.71 (apparent t, J = 10 Hz, 1H), 4.59 (dd, J = 10.9 and 4.0 Hz, 1H), 4.08 (s, 3H), 4.07-3.88 (m, 2H), 3.98 (s, 3H), 3.95 (s, 3H), 3.92 (s, 3H), 3.61 (dd, J = 11 and 9.4 Hz, 1H); ¹³C NMR (CDCl₃, 63 MHz) δ 159.9 (C), 159.3 (C), 150.1 (C), 146.7 (C), 144.7 (C), 143.1 (C), 141.0 (C), 140.5 (C), 138.8 (C), 136.0 (C), 129.6 (C), 128.5 (CH), 128.1 (CH), 127.7 (CH), 125.4 (C), 123.9 (C), 123.5 (C), 114.7 (C), 111.5 (CH), 106.4 (CH), 102.4 (CH), 97.6 (CH), 70.9 (CH₂), 61.4 (CH₃), 61.0 (CH₃), 56.2 (CH₃), 55.0 (CH₂), 52.3 (CH₃), 46.3 (CH₂), 43.0 (CH); IR (KBr) v_{max} 3459, 2936, 1732, 1622, 1494, 1308, 1203, 744, 697 cm⁻¹; UV (EtOH) λ_{max} 208.5 (ϵ = 35611), 297 (ϵ = 22216), 326 $(\epsilon = 24475)$ nm; FDMS *m*/*z* 604 (M⁺, 100). Anal. Calcd for C32H29N2O8Cl: C, 63.52; H, 4.83; N, 4.63; Cl, 5.86. Found: C, 63.58; H, 4.87; N, 4.39; Cl, 6.02.

(-)-(1R)-Methyl 5-(Benzyloxy)-1-(chloromethyl)-1,2-dihydro-3-[(5,6,7-trimethoxy-1H-indol-2-yl)carbonyl]-3Hfurano[3,2-e]indole-7-carboxylate (17b). Amine 15b (216 mg, 0.530 mmol) was coupled to 5,6,7-trimethoxyindolecarboxylic acid 16, in the same manner described above, to furnish amide 17b (161 mg, 50%, HPLC (analytical ChiraCel OD column; flow rate = 1.0 mL/min; UV detection at λ = 254 nm; eluent, CH₃CN:H₂O 90:10): **17b:**: $t_R = 8.42$ min; 94 % ee) as a white solid: $[\alpha]^{23}_D - 13.2^{\circ}$ (*c* 0.5, CHCl₃); mp 138–140 °C; ¹H NMR (CDCl₃, 300 MHz) δ 9.41 (brs, 1H), 8.31 (s, 1H), 7.55– 7.51 (m, 3H), 7.43-7.33 (m, 3H), 6.95 (d, J = 2.1 Hz, 1H), 6.86 (s, 1H), 5.35 (s, 2H), 4.70 (apparent t, J = 9.9 Hz, 1H), 4.58 (dd, J = 10.9 and 3.9 Hz, 1H), 4.08 (s, 3H), 4.04-3.88 (m, 2H),3.98 (s, 3H), 3.94 (s, 3H), 3.91 (s, 3H), 3.60 (dd, J = 10.8 and 9.6 Hz, 1H); 13 C NMR (CDCl₃, 63 MHz) δ 159.9 (C), 159.3 (C), 150.1 (C), 146.7 (C), 144.7 (C), 143.1 (C), 141.0 (C), 140.5 (C), 138.8 (C), 136.0 (C), 129.6 (C), 128.5 (CH), 128.1 (CH), 127.7 (CH), 125.4 (C), 123.9 (C), 123.5 (C), 114.7 (C), 111.5 (CH), 106.4 (CH), 102.4 (CH), 97.6 (CH), 70.9 (CH2), 61.4 (CH3), 61.0 (CH₃), 56.2 (CH₃), 55.0 (CH₂), 52.3 (CH₃), 46.3 (CH₂), 43.0 (CH); IR (KBr) v_{max} 2937, 1731, 1622, 1494, 1416, 1309, 1204, 1109, 744, 697 cm⁻¹; UV (EtOH) $\lambda_{\rm max}$ 209 (ϵ = 44076), 297.5 (ϵ = 26058), 326 (ϵ = 30391) nm; FDMS *m*/*z* 603.7 (M⁺, 100). Anal. Calcd for C₃₂H₂₉N₂O₈Cl: C, 63.53; H, 4.83; N, 4.63; Cl, 5.86. Found: C, 63.74; H, 4.81; N, 4.40; Cl, 6.01.

Methyl (1S)-5-hydroxy-1-(chloromethyl)-1,2-dihydro-3-[(5,6,7-trimethoxy-1H-indol-2-yl)carbonyl]-3H-furano-[3,2-e]indole-7-carboxylate (3a). To a solution of benzyl ether 17a (149 mg, 0.247 mmol) in tetrahydrofuran (5.80 mL) was added 10% aqueous ammonium formate (560 μ L). The solution was cooled to 0 °C and 10% Pd-C (56 mg) added. The reaction mixture was stirred for 5 h, filtered through a pad of Celite, and concentrated to provide (+)-oxaduocarmycin SA (**3a**) (122 mg, 96%) as a yellow powder: $[\alpha]^{23}_{D} + 1.73^{\circ}$ (*c* 0.11, DMF); mp 265–268 °C dec; ¹H NMR (DMSO- d_6 , 300 MHz) δ 11.39 (d, J = 1.14 Hz, 1H), 10.52 (s, 1H), 7.96 (s, 1H), 7.92 (s, 1H), 6.97 (d, J = 1.7 Hz, 1H), 6.92 (s, 1H), 4.66 (apparent t, J = 10 Hz, 1H), 4.34 (dd, J = 10.6 and 3.54 Hz, 1H), 4.08-4.02 (m,1H), 3.98-3.94 (m, 1H), 3.88 (s, 3H), 3.87 (s, 3H), 3.77 (s, 3H), 3.75 (s, 3H), 3.34-3.26 (buried m, 1H); ¹³C NMR (DMSOd₆, 63 MHz) δ 160.0 (C), 159.1 (C), 149.2 (C), 145.6 (C), 142.6 (C), 141.9 (C), 140.9 (C), 139.9 (C), 139.1 (C), 131.0 (C), 125.3 (C), 124.2 (C), 123.2 (C), 114.3 (C), 113.0 (CH), 106.0 (CH), 104.0 (CH), 98.1 (CH), 61.1 (CH₃), 61.0 (CH₃), 56.0 (CH₃), 54.8 (CH₂) 52.3 (CH₃), 47.4 (CH₂), 41.4 (CH); IR (KBr) v_{max} 3449,

1726, 1589, 1492, 1453, 1310, 1198, 1106 cm⁻¹; UV (EtOH) λ_{max} 304 (ϵ = 8970), 328 (ϵ = 9326) nm; FDMS *m*/*z* 514 (M⁺, 100). Anal. Calcd for C₂₅H₂₃N₂O₈Cl: C, 58.31; H, 4.50; N, 5.44; Cl, 6.89. Found: C, 58.25; H, 4.66; N, 5.29; Cl, 6.61.

Methyl (1R)-5-Hydroxy-1-(chloromethyl)-1,2-dihydro-3-[(5,6,7-trimethoxy-1H-indol-2-yl)carbonyl]-3H-furano-[3,2-e]indole-7-carboxylate (3b). Benzyl ether 17b (130 mg, 0.215 mmol) was deprotected, in the same manner described previously, to give (–)-oxaduocarmycin SA (**3b**) (109 mg, 99%) as a yellow solid: $[\alpha]^{23}_D - 2.1^\circ$ (*c* 0.12, DMF); mp 264–267 °C; ¹H NMR (DMSO- d_6) δ 11.40 (s, 1H), 10.51 (s, 1H), 7.96 (s, 1H), 7.94 (s, 1H), 6.98 (d, J = 1.7 Hz, 1H), 6.92 (s, 1H), 4.67 (apparent t, J = 10 Hz, 1H), 4.32 (dd, J = 11.4 and 4.2 Hz, 1H), 4.08-4.05 (m, 1H), 3.98-3.92 (m, 1H), 3.89 (s, 3H), 3.87 (s, 3H), 3.77 (s, 3H), 3.75 (s, 3H), 3.36-3.27 (buried m, 1H); ¹³C NMR (DMSO-*d*₆, 63 MHz) δ 160.0 (C), 159.1 (C), 149.2 (C), 145.6 (C), 142.6 (C), 141.9 (C), 140.9 (C), 139.9 (C), 139.1 (C), 131.0 (C), 125.3 (C), 124.2 (C), 123.2 (C), 114.3 (C), 113.0 (CH), 106.0 (CH), 104.0 (CH), 98.1 (CH), 61.1 (CH₃), 61.0 (CH₃), 56.0 (CH₃), 54.8 (CH₂) 52.3 (CH₃), 47.4 (CH₂), 41.4 (CH); IR (KBr) $v_{\rm max}$ 3450, 1725, 1586, 1492, 1429, 1310, 1107, 746 cm⁻¹; UV (EtOH-sparingly soluble) λ_{max} 208, 303, 328 nm; FDMS m/z514 (M⁺, 100). Anal. Calcd for C₂₅H₂₃N₂O₈Cl: C, 58.31; H, 4.50; N, 5.44; Cl, 6.89. Found: C, 58.55; H, 4.67; N, 5.14; Cl, 6.92

In Vitro Cytotoxicity Assays. T222 (human lung epidermoid carcinoma)¹⁴ cells (1 × 10⁴) were distributed in each well of 96 well tissue culture plates and incubated in leucinedeficient media (leucine-free DMEM, 13 µg/mL L-leucine, 29.9 µg/mL L-glutamine, 50 µg/mL gentamicin, and 10% dialyzed fetal bovine serum) for 16 h at 37 °C in 5% carbon dioxide/air atmosphere. The medium was removed aseptically and compound dilutions added in leucine-deficient medium (200 µL). After 48 h the media was removed and 4 µCi (³H-Leucine-NEN, Boston, MA) was added to each well. The plates were returned to the incubator for 24 h. Radioactivity incorporated into macromolecules was detemined using an automated cell harvester and liquid scintillation techniques. Data were evaluated as % reduction in incorporation of radioactivity relative to controls incubated in medium without compound to yield a 50% cytotoxic concentration (IC₅₀).

Å modification of the original MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) colorimeteric assay, described by Mosmann³³ that measures the reduction of MTT to a violet-colored formazan by living cells, was used for the CCRF-CEM and GC3/C1 cell assays. The tumor cells (1 \times 10⁴) were seeded in assay medium (100 μ L)/well in 96-well flat bottom tissue culture plates (Costar, Cambridge, MA). Assay medium consisted of RPMI-1640 medium supplemented with 10% dialyzed fetal bovine serum. Well 1A was left blank (100 μ L of growth medium without cells). Stock solutions of test compounds were prepared in DMSO at 1 mg/mL, and a series of 2-fold dilutions were made in Dulbecco's phosphate-buffered saline (PBS). Aliquots (10 μ L) of each concentration were added to triplicate wells. The compounds were tested at concentrations ranging from 0.01 to 0.00008 μ g/mL. Plates were incubated for 72 h at 37 °C in a humidified atmosphere of 5% CO2-in-air. MTT was dissolved in PBS at 5 mg/mL, and following incubation of plates, a stock solution of MTT (10 μ L) was added to the assay wells and the plates were incubated at 37 °C for an additional 2 h. DMSO (100 µL) was added to each well to thoroughly solubilize the formazan, and the plates were then read on a Dynatech (Alexandria, VA) MR600 reader using a test wavelength of 570 nm and a reference wavelength of 630 nm. The IC₅₀ was determined as the concentration of drug required to inhibit cell growth by 50%.

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